6-[¹⁸F]Fluorodopamine 6-[¹⁸F]FDA

Created: April 06, 2005 Updated: April 18, 2005

Chemical name: 6-[¹⁸F]Fluorodopamine **Abbreviated name:** 6-[¹⁸F]FDA, [¹⁸F]FDA

Synonym:

Backbone: Compound

Target: Sympathetic neuron transporters

and dopamine β-hydroxylase

Mechanism: Uptake, conversion to

norepinephrine and vesicle

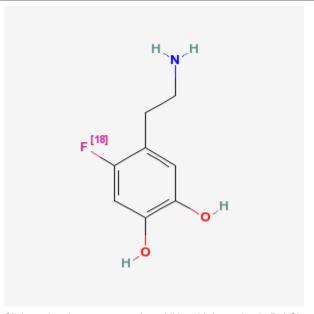
storage

Method of detection: PET
Source of signal: 18F
Activation: No
In vitro studies: Yes
Rodent studies: Yes
Other non-primate mammal
Studies: Studies:

studies:

Non-human primate studies: Yes

Human studies: Yes



Click on the above structure for additional information in PubChem [http://pubchem.ncbi.nlm.nih.gov/].

Background

[PubMed]

Dopamine, a neurotransmitter, plays an important role in the mediation of movement, cognition, and emotion. Parkinson's disease (PD) is associated with a loss of dopamine-containing neurons in the striatum of the brain (1, 2). Dopamine is synthesized within nerve cells (3). Chemically, L-tyrosine is converted to dihydroxyphenylalanine (L-DOPA) and then to dopamine in a two-step process. The first rate-limiting step is catalyzed by tyrosine 3-monoxygenase (tyrosine hydroxylase or TH). The second step is catalyzed by aromatic L-amino acid decarboxylase (L-DOPA decarboxylase, AAAD). In dopaminergic neurons, dopamine is not metabolized further and is stored in vesicles in the presynaptic nerve terminals. Interstitial dopamine is recaptured by the dopamine transporter, DAT.

In noradrenergic neurons, dopamine is converted to norepinephrine (NE) by dopamine β -hydroxylase and stored in vesicles in the neurons (4). Released NE in synaptic junctions is either inactivated by COMT in postsynaptic cells or transported by a NE transporter (NET) into the nerve

terminals (uptake-1). Dopamine is also efficiently transported by NET. At extraneuronal locations, DAT is present in the placenta and lung endothelial cells, and NET is present in the stomach and pancreas. There are also three non-neuronal transporters functioning in peripheral tissues such as the heart, liver, kidneys, intestine, blood vessels, retina, and placenta. These uptakes by non-neuronal cells are termed uptake-2.

After transported into sympathetic nerve endings by uptake-1, 6-[¹⁸F]FDA is rapidly converted to 6-[¹⁸F]fluoronorepinephrine (6-FNE) by dopamine β-hydroxylase in neuronal vesicles (5). 6-[¹⁸F] FDA is also metabolized via mitochondrial monoamine oxidase to yield [¹⁸F]6-fluoro-3,4-dihydroxyphenylacetic acid (FDOPAC). In nonneuronal cells, 6-[¹⁸F]FDA is converted by COMT sequentially to *O*-[¹⁸F]methoxytyramine and [¹⁸F]6-fluorochomovanillic acid (FHVA). FDOPAC taken up after release from sympathetic neurons by uptake-2 is converted to FHVA by COMT. Uptake of 6-[¹⁸F] FDA into sympathetic nerve terminals, with conversion to and storage of 6-[¹⁸F]FNE in vesicles, would lead to more intense positron emission tomography (PET) signals from sympathetically innervated tissues than non-innervated tissues.

Synthesis

[PubMed]

 $6-[^{18}F]FDA$ was synthesized from $6-[^{18}F]FDOPA$ by enzymatic decarboxylation using AAAD prepared from hog kidneys with a radiochemical purity of >98% and a specific activity of 37 MBq/mg (1 mCi/mg) (5). A direct synthesis was also used to produce a higher specific activity. *N*-(Trifluoroacetyl)-3,4-dimethoxy-6-trifluoroacetoxymercuro- β -phenethyamine was fluorodemercurated by $[^{18}F]F_2$ to form $6-[^{18}F]FDA$ with a radiochemical purity of 98% and about 22 MBq/mmol (800 mCi/mmol) at the end of synthesis (6).

To obtain a higher specific activity, a multi-step synthesis of 6-[¹⁸F]FDA using nucleophilic aromatic substitution by [¹⁸F]fluoride ion/Kryptofix2.2.2 was achieved in 105 min with a radiochemical yield of 20% and a specific activity of 74-185 GBq/μmol (2-5 Ci/μmol) at the end of bombardment (7). Chemical and radiochemical purities were >98%.

6-[18 F]FDA was routinely synthesized by the direct fluorination of dopamine with [18 F]KF using the standard [18 F] potassium Kryptofix complex (8). Reverse-phase high-performance liquid chromatography was used to separate 6-[18 F]FDA from the reaction mixture containing 2- and 5-[18 F]FDA. The radiochemical yield of 6-[18 F]FDA was $10 \pm 2\%$ at the end of the 120-min synthesis from the end of bombardment. The specific activity of 6-[18 F]FDA was 370 GBq/mmol (10 Ci/mmol) at the end of synthesis.

In Vitro Studies: Testing in Cells and Tissues

[PubMed]

The enzyme kinetic parameters using Lineweaver-Burk plots for uptake of [3 H]DA and 6-[18 F] FDA were determined *in vitro* from bovine chromaffin granule membranes (9). The K_m and V_{max} for [3 H]DA were 14.7 μ M and 1.6 nmol/mg protein-min, respectively. The K_m and V_{max} for 6-[18 F]FDA

were 15.3 μ M and 2.4 nmol/mg protein-min, respectively. FDA showed an affinity comparable to DA. Both uptakes were inhibited by reserpine, an uptake-1 blocker. Unlabeled FDA inhibited uptake of [3 H]DA with a K_{i} of 20.8 μ M.

Animal Studies

Rodents

[PubMed]

A high uptake of radioactivity was found in the kidneys, left ventricular myocardium, spleen, salivary gland, and liver of rats at 5 min after injection of [³H]6-FDA (10). The amount of radioactivity was decreased to <3% in plasma in 5 min. Tissue/blood ratios increased progressively from 5 to 120 min. In the heart, the majority of the tissue radioactivity was [³H]6-FDA (17%) and [³H]6-FNE (70%). In the kidneys and liver, [³H]FDA and [³H]FNE contributions were <30%. No [³H]6-FNE was detected in the plasma.

The neuronal uptake and metabolism of $6 \cdot [^{18}F]FDA$ and $[^{3}H]dopamine$ were studied in the heart, submaxillary gland, and spleen. Less $6 \cdot [^{18}F]FNE$ accumulation in tissues than $[^{3}H]NE$ was observed as a result of inefficient hydroxylation of fluorinated dopamine (11). $6 \cdot [^{18}F]FDA$ was rapidly stored in vesicles of sympathetic neurons but was a poorer substrate than $[^{3}H]DA$ to form NE. Accumulation of $[^{3}H]$ catechols in the heart was decreased by $64 \cdot 88\%$ when unlabeled $6 \cdot FDA$ ($250 \mu g/kg$) was coinjected with a trace amount of $[^{3}H]DA$. Desipramine and reserpine (uptake-1 blockers) pretreatment of rats blocked the tissue accumulation of tritiated and fluorinated dopamine and their dihydroxymetabolites.

Other Non-Primate Mammals

[PubMed]

In dogs, 6-[¹⁸F]FDA PET images were intense in the renal pelvis, heart, liver, kidneys, spleen, and salivary glands, with little signal from the brain, lungs, and skeletal muscle at 1 h after injection (10). Concentrations of radioactivity in blood and plasma fell rapidly with a biologic half-life of 1.5 min. The left ventricular myocardium was strikingly delineated. High radioactivity in the gallbladder and urinary system pointed to the hepatic and renal excretion of the tracer and its metabolites. Treatment with 6-hydroxydopamine (6-OHDA, neurotoxin) or reserpine (uptake-1 blocker) diminished almost completely the signal from the left ventricular myocardium (12). Plasma FDOPAC levels were lowered in 6-OHDA treated dogs and elevated in reserpinized dogs. Decreased PET signal from denervated salivary gland also confirmed that 6-[¹⁸F]FDA was accumulated normally in sympathetic neurons (5).

Non-Human Primates

[PubMed]

Comparative PET studies of (-)- and (+)-6-[18F]FNE and of 6-[¹⁸F]FDA in the heart were performed in the same baboon (13). There was a longer retention of radioactivity after injection of (-) FNE than for the (+)FNE. There was an initially higher and faster uptake in the heart for 6-[¹⁸F]FDA than for FNE. All three tracers disappeared rapidly in plasma. There was a greater washout also for 6-[¹⁸F]FDA than for FNE. There was only 1-2% intact FDA as compared with 28% for (-)FNE and 17% for (+)FNE at 10 min after injections. Most of the metabolites were methylated non-catechols. Metabolites appeared more rapidly for FDA than (-)FNE and (+)FNE. Desipramine (a specific NET blocker; 0.5 mg/kg) blocked almost completely the uptake of (-)FNE, and there was a 60-70% recovery of FNE uptake at 24 h after desipramine pretreatment. The uptake of 6-[¹⁸F]FDA was only partially blocked by desipramine at the same dosage, suggesting additional reuptake mechanisms for 6-[¹⁸F]FDA accumulation in the heart.

Human Studies

[PubMed]

Human dosimetry was estimated in 10 healthy volunteers (14). The bladder wall receives the highest dose (0.22 mGy/MBq or 0.83 rad/mCi). Other organs receiving high doses are the kidneys (0.19 mGy/MBq or 0.71 rad/mCi) and spleen (0.037 mGy/MBq or 0.14 rad/mCi). The liver and lungs receive <0.016 mGy/MBq (0.06 rad/mCi). The effective dose equivalent of 0.0068 mSv/MBq (25 mrem/mCi) was estimated in the intravenous administration of 6-[¹⁸F]FDA.

6-[¹⁸F]FDA PET scans were obtained in healthy volunteers after intravenous injection of 37-148 MBq (1-4 mCi) 6-[¹⁸F]FDA (15). The left ventricular myocardium was visualized in all subjects. Arterial plasma FDA concentration decreased rapidly with a biologic half-life of 2.4 min. FHVA and FDA sulfate appeared rapidly in the plasma. No FNE and its metabolites were detected. Greater than 94% of FDA was excreted mostly as metabolites in urine at 24 h after injection. Desipramine (uptake-1 blocker) decreased the uptake of myocardial radioactivity and plasma FDOPAC. Trimethaphan (TRI), a postganglion nerve traffic blocker, induced higher levels of 6-[¹⁸F]FDA PET myocardial signal than untreated subjects. TRI increased FDOPAC plasma levels but decreased FNE levels as compared with untreated subjects. Tyramine (TYR) displaces amines from vesicles and induces release of NE into the synaptic cleft. TYR decreased the 6-[¹⁸F]FDA PET myocardial signal more rapidly than in untreated subjects. TYR increased plasma levels of FNE and its metabolite by 30% and 78%, respectively. FDA PET scanning is a useful tool to assess sympathetic innervation and function noninvasively in human myocardium.

6-[¹⁸F]FDA PET permits objective monitoring of cardiac sympathetic innervation and function in various disease conditions, such as Parkinson's disease (16), hypertension, and congestive heart failure (17). In recent studies, 6-[¹⁸F]FDA has also demonstrated its usefulness in the imaging of chromaffin tumors, neuroblastomas, ganglioneuromas, and metastatic pheochromocytomas (18, 19).

References

- 1. Carbon M, Ghilardi MF, Feigin A, Fukuda M, Silvestri G, Mentis MJ, Ghez C, Moeller JR, Eidelberg D. Learning networks in health and Parkinson's disease: reproducibility and treatment effects. Hum Brain Mapp 19:197–211; 2003. (PubMed)
- Chesselet MF, Delfs JM. Basal ganglia and movement disorders: an update. Trends Neurosci 19:417–422; 1996. (PubMed)
- 3. Barrio JR, Huang SC, Phelps ME. Biological imaging and the molecular basis of dopaminergic diseases. Biochem Pharmacol 54:341–348; 1997. (PubMed)
- 4. Eisenhofer G. The role of neuronal and extraneuronal plasma membrane transporters in the inactivation of peripheral catecholamines. Pharmacol Ther 91:35–62; 2001. (PubMed)
- 5. Goldstein DS, Chang PC, Eisenhofer G, Miletich R, Finn R, Bacher J, Kirk KL, Bacharach S, Kopin IJ. Positron emission tomographic imaging of cardiac sympathetic innervation and function. Circulation 81:1606–1621; 1990. (PubMed)
- Goldstein DS, Eisenhofer G, Dunn BB, Armando I, Lenders J, Grossman E, Holmes C, Kirk KL, Bacharach S, Adams R. Positron emission tomographic imaging of cardiac sympathetic innervation using 6-[18F] fluorodopamine: initial findings in humans. J Am Coll Cardiol 22:1961–1971; 1993. (PubMed)
- Ding YS, Fowler JS, Gatley SJ, Dewey SL, Wolf AP, Schlyer DJ. Synthesis of high specific activity 6-[18F] fluorodopamine for positron emission tomography studies of sympathetic nervous tissue. J Med Chem 34:861–863; 1991. (PubMed)
- 8. Chirakal R, Coates G, Firnau G, Schrobilgen GJ, Nahmias C. Direct radiofluorination of dopamine: 18F-labeled 6-fluorodopamine for imaging cardiac sympathetic innervation in humans using positron emission tomography. Nucl Med Biol 23:41–45; 1996. (PubMed)
- 9. Endres CJ, Swaminathan S, DeJesus OT, Sievert M, Ruoho AE, Murali D, Rommelfanger SG, Holden JE. Affinities of dopamine analogs for monoamine granular and plasma membrane transporters: implications for PET dopamine studies. Life Sci 60:2399–2406; 1997. (PubMed)
- Goldstein DS, Chang PC, Smith CB, Herscovitch P, Austin SM, Eisenhofer G, Kopin IJ. Dosimetric estimates for clinical positron emission tomographic scanning after injection of [18F]-6-fluorodopamine. J Nucl Med 32:102–110; 1991. (PubMed)
- 11. Eisenhofer G, Hovevey-Sion D, Kopin IJ, Miletich R, Kirk KL, Finn R, Goldstein DS. Neuronal uptake and metabolism of 2- and 6-fluorodopamine: false neurotransmitters for positron emission tomographic imaging of sympathetically innervated tissues. J Pharmacol Exp Ther 248:419–427; 1989. (PubMed)
- 12. Goldstein DS, Grossman E, Tamrat M, Chang PC, Eisenhofer G, Bacher J, Kirk KL, Bacharach S, Kopin IJ. Positron emission imaging of cardiac sympathetic innervation and function using 18F-6-fluorodopamine: effects of chemical sympathectomy by 6-hydroxydopamine. J Hypertens 9:417–423; 1991. (PubMed)
- 13. Ding YS, Fowler JS, Dewey SL, Logan J, Schlyer DJ, Gatley SJ, Volkow ND, King PT, Wolf AP. Comparison of high specific activity (-) and (+)-6-[18F]fluoronorepinephrine and 6-[18F]fluorodopamine in baboons: heart uptake, metabolism and the effect of desipramine. J Nucl Med 34:619–629; 1993. (PubMed)
- 14. Goldstein DS, Coronado L, Kopin IJ. 6-[Fluorine-18]fluorodopamine pharmacokinetics and dosimetry in humans. J Nucl Med 35:964–973; 1994. (PubMed)
- 15. Goldstein DS, Holmes C, Stuhlmuller JE, Lenders JW, Kopin IJ. 6-[18F]fluorodopamine positron emission tomographic scanning in the assessment of cardiac sympathoneural function--studies in normal humans. Clin Auton Res 7:17–29; 1997. (PubMed)
- 16. Li ST, Dendi R, Holmes C, Goldstein DS. Progressive loss of cardiac sympathetic innervation in Parkinson's disease. Ann Neurol 52:220–223; 2002. (PubMed)
- 17. Langer O, Halldin C. PET and SPET tracers for mapping the cardiac nervous system. Eur J Nucl Med Mol Imaging 29:416–434; 2002. (PubMed)
- 18. Ilias I, Shulkin B, Pacak K. New functional imaging modalities for chromaffin tumors, neuroblastomas and ganglioneuromas. Trends Endocrinol Metab 16:66–72; 2005. (PubMed)

19. Goldstein DS, Eisenhofer G, Flynn JA, Wand G, Pacak K. Diagnosis and localization of pheochromocytoma. Hypertension 43:907–910; 2004. (PubMed)